

MODELING AND OPTIMIZATION OF CEPHALOSPORIN C PRODUCTION USING
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ABSTRACT: The objective of this study is to improve the cephalosporin C production through optimization of medium and culture conditions. The effect of agitation speed on CPC production in 3 l bioreactor was investigated with fed batch mode. The maximum CPC (0.79 g/l), the biomass (35.4g) was obtained at 400 rpm. A statistical method was introduced to optimize the main culture medium constituents. Some medium constituents involving glucose, ammonium sulphate as inorganic nitrogen source and methionine were found to be the most effective factors for (CPC) production. The results showed the priority of fed batch culture than the batch one, since it increased the yield of the CPC by about 155 %.

Key words; Cephalosporin C (CPC), Production, *Streptomyces*

INTRODUCTION

Cephalosporin together with penicillin belong to the family of β -lactam antibiotics, cephalosporins are important broad spectrum antibiotics in the international market and are considerably more resistant to β -lactamases than penicillin. A number of reports are available on the various aspects of CPC production such as production in batch as well as in continuous mode using free and immobilized cells of molds (Srivastava and Kundu, 1999; Cruz *et al.*, 2004, and Kim *et al.*, 2005). The biosynthesis of CPC is studied in bioreactor using synthetic medium, it was observed that with the progress of fermentation process increase in the viscosity of the broth occurs and thereby decreasing oxygen mass transfer (Kundu *et al.*, 2003, Verma *et al.*, 2006). The present investigation deals with the optimization of medium constituents to increase the CPC productivity using the strain of *Streptomyces*.

MATERIALS AND METHODS

Microorganism

The production of CPC was carried out by *Streptomyces gresiolus* obtained from Genetic and cytology Department, National Research center, Dokki, Cairo, Egypt.

Cultivation conditions

The stock culture was maintained by transferring the organism monthly on potato dextrose agar slants. The basal production medium consists of 2.5 % glucose, 0.4% (NH₄)₂ SO₄, 0.3 KH₂PO₄ and 0.5DL, methionine. Sugar and ammonium sulphate were sterilized separately from other components. Erlenmeyer flasks containing 50ml medium, the culture was maintained at 300 rpm at 27°C on a rotary shaking incubator for inoculum preparation. The fed batch fermentation in the stirred tank fermentor (3l) was carried out at 27°C and the operating volume 2 liter with flow rate 1 vvm and 5 % (v/v) of glucose was fed at 3rd and 5th day into the bioreactor culture broth according to the method described by Srivastava *et al.*, (1999).

Analytical methods

The dry cell weight of mycelium measured as follow: 10 ml of culture broth was centrifuged at 12000xg and filtered through a pre-weighted what man glass microfiber filter GF/C, after being dried it washed twice with distilled water. The cells were dried at 95 C to a constant weight prior to measured by using the method described by Miller , 1959 and Matsumura *et al.*, 1981).

CPC was measured firstly using bioassay technique where *bacillus subtilis* used as a test microorganism . It was also measured by high liquid chromatography (HPLC) using reverse phase column of u band a pak C₁₈ and 254nm UV detector . the mobile phase was acetonitrile phosphate buffer, the elution mixture was 98 % phosphate buffer and 2 % acetonitrile with a flow rate 0.9ml'min. cephalosporin C zinc salts obtained from Sigma USA was used as standard.(Cruz, *et al.*, 2004).

Experimental Design

Factional factorial design was employed to optimized the main medium constituents using analysis of variance (ANOVA) and the optimal concentration of the main medium contents which had a significant effect on CpC production was determined . the variable was coded according to the equation.

$$X_i = (X_i - X_0) / \Delta X \quad i=1,2,3,4,\dots\dots\dots$$

Where

X_i is the coded (dimensionless) value of the variable

X₁, X₀ the value of X_i at the center point , ΔX the step change

RESULTS AND DISCUSSION

Effect of different fermentation time on the Production of CPC using shake flask

The results in table (1) showed that time course of CPC production in shake flask using an optimized main medium. The maximum CPC production (0.46 g/l) was obtained at 5 day and then CPC production was gradually decreased. The maximum CPC production was similar to that predicted by statistical analysis .but as glucose deleted, cell mass and CPC production gradually decreased .form these results it was thought that the carbon source is necessary to maintenance of the activity and productivity process.

Table-1: Effect of different fermentation time on the production of CPC using shake flask

Fermentation time (day)	Biomass (g/l)	CPC production(g/l)
2	1.72	0.22
3	2.31	0.31
4	2.50	0.36
5	2.41	0.46
6	2.62	0.30
7	2.60	0.25

Effect of different agitation speed on CPC production using laboratory stirred ferment or

In the present investigation the effect of different agitation speed (200, 300, 400, and 500 rpm) were tested for the production of CPC process. The results presented in t able (2) and showed that the best CPC out put (0.79mg/L) was obtained at 400 rpm .while the other speeds showed reduced levels of CPC .

Table (2): Effect of different agitation speed on the production of CPC

Agitation speed (rpm)	Biomass (g/L)	CPC production (g/L)
200	24.64	0.54
300	31.52	0.62
400	35.41	0.79
500	29.2	0.55

Optimization of the main medium constituents using statistical methods

From preliminary experiments made in order to examine the effects of different components on CPC production. Three factors which play the most important role in the synthesis process. These were glucose, ammonium sulphate and DL methionine . Tables (3,4) showed the value of these variables at different levels . The CPC production varied in a range of 0.047 g/l . on the basis of thesis experimental values statistical testing was carried out using analysis of variance (ANOVA) as shown in Table (5) , when the test was applied an each factor for CPC production with three variable .

Table 3: Level of the variable tested in functional factorial design

Medium constituents	Symbol	Coded value		
		-1	0	1
glucose	A	1.2	2.9	4.2
Ammonium sulphate	B	0.4	0.8	1.1
DL methionine	D	0.3	0.5	0.8

The behavior of the system was explained by the following second degree polynomial equation

$$Y = B_0 + S B_i x_i + S B_{ii} + i_2 + S B_{iii} + i_3 + \dots$$

Where, y is the predicted response. B_0 the offset best term

B_i the liner effect, B_{ii} the square effect and B_i the interaction effect

Table (4) Experimental design and results for analysis of variance (ANOVA)

Run	A	B	C	CPC production
1	-	-	-	0.420
2	-	-	-	0.449
3	+	-	-	0.350
4	+	-	-	0.375
5	-	+	-	0.185
6	-	+	-	0
7	+-	+	-	0
8	-	+	+	0.478
9	-	-	+	0.349
10	+	-	+	0.378
11	+-	-	+	0
12	-	-	+	0
13	-	-	+	0.41
14	+	+	+	0.372
15	+	+	+	0.310

Table (5) Satirical analysis of factors through analysis of variance

Factor	CPC production	
	F value	P value
A	867.60	0.0001
B	713.01	0.0001
C	41.05	0.1114
AB	121.5	0.2227
AC	187.83	0.0006
BC	15.48	0.0300

Table (6): Experimental design and results for determination of optimal concentration of glucose and ammonium sulphate

Run	Glucose	Ammonium sulphate	CPC production
1	-1	-1	0.270
2	1	-1	0
3	-1	1	0.185
4		1	0.315
5	1.314	0	0
6	0	1.414	0
7	1.414	0	0
8	0	1.414	0.334
9	0	0	0.340
10	0	0	0.408

Glucose and ammonium sulphate were statistically at 1% level of significant, so the effect of each component on CPC production was investigated. Especially, in the case of glucose the F and P value for CPC were 867.60 and 0.0001 respectively. From these results, it was concluded that glucose was the most effective components on CPC production.

The basis of the experimental design given in table (6) using these results, the following second order polynomial equation relating to the CPC production explain the experimental data.

$$\text{CPC production} = 0.478 - 0.005X - 0.188X^2 + 0.101y - 0.14Y^2 + 0.076XY$$

Where X = glucose concentration

Y = ammonium sulphate concentration

CONCLUSION

The results obtained showed that the productivity of CPC was affected by different factors (glucose, ammonium sulphate, agitation speed), it revealed that glucose concentration was highly significant on the production process than the other parameters tested.

REFERENCES

- Behman ,T; Ehan, G.; Termeh, T.; Elnaz,P.and Mahdi, R.C.(2012).Comparative evaluation of cephalosporin C production in solid state fermentation and submerged liquid culture . J. Microbiol. Biotechnol. and food sciences 2 (1) pp 83-94.
- Cruz. A.;Pan T.; Giordano, R., C.; and Araujo M., L; (2004). Cephalosporin C production by immobilized *Cephalosporium acremonium* cells in repeated batch tower bioreactor . Biotechnol. Bioeng.,85 pp (96-102).
- Duan .; Yan , G; Zhao,y.;Li,H; Ni,w; Sang,M.; Liu,L.; and Shi, Z.; (2012) Enhanced cephalosporin C production with a combinational ammonium sulphate and do state based soy bean oil feeding strategy . Biochemical Engineering journal vol. (61) pp-1-10.
- Matsumura, E., , Swartz, R., and Zan, C . (1981). Pencillins biosynthetic and semi-synthetic in economic microbiology . Secondary products of metabolism .edited by AH Rose (Academic press, London. 35- 120.
- Miller , G., L.; (1959) . Use of dinitro-salicylic acid reagent for determination of reducing sugar . Analytical chemistry , 31, 426- 428.
- Paul .L.,S.; Anthony , j.; Thomas, D ;Cathleen, A ; fisher , J.; and Stephen , W .; (1989). Use of Recombinant DNA to improve production of cephalosporin C by *Cephalosporium acremonium* . Biotechnology (7),pp- 477-485.
- Rui-juan Bin; qioa, Bing- zhi li ;Hualu, Yao chen and Ying –jin Yuan (2012). Comparative lipodomic analysis of *cephalosporium acremonium* in sight into industrial and pilot fermentation. JSIR vol. (17) n0 .2 pp- 259-269 .
- Srivastava , P., and Kundu , S.(1998) . A comparative evaluation of Cephalosporin C production using various immobilized modes . J ., Gen. Appl. Microbiol. 44 pp(113-117.
- Srivastava , P.; Mishra, P.;and Kindu, S.(2006). Process strategies for Cephalosporin C fermentation. JSIR vol., 65(07) July 2006
- Weichang, Z.; Karin Holzhaure ,R.; Thomas, B. and Kari, S.(1993).Cephalosporin C production by a highly productive *Cephalosporium acremonium* strain an air lift tower loop reactor with static mixers. Natur biotechnology11, 926- 929.