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## TOTAL AND PROTEIN-BOUND SIALIC ACID SERUM LEVELS IN PREDIABETES

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#### Abstract

Background: Prediabetes is the main risk factor of Type 2 diabetes mellitus (T2DM), which is a low-grade systemic chronic inflammation disease. Sialic acid is a marker of systemic inflammation as well as significantly associated with new T2DM. Previous studies are scarce and have yielded contrasting results about sialic acid levels in prediabetes. The aim of this study was to assess sialic acid serum levels (total sialic acid and its fractions) innewly diagnosed prediabetes. Materials and Methods: Potential confounding variables were controlledto search for the association of newly diagnosed prediabetes with total sialic acid (TSA), protein-bound sialic acid (BSA), and free sialic acid (FSA) serum levels; 28 cases of adults with prediabetes and 31 normoglycemic controls, matched for BMI, sex and age were included. Results: Compared to controls, only TSA and BSA were elevated in subjects with prediabetes ( $p<0.0001$ and $p$ $<0.0001$, respectively). A positive association between prediabetes and both, TSA and BSA, was observed ( $\beta=11.9, \mathrm{p}$ $<0.0001$ and $\beta=12.0, p<0.0001$, respectively). Conclusions: Our results showed increased TSA and BSA levels in adults with newly diagnosed prediabetes. Increased TSA relied on the increment of BSA, and both, TSA and BSA were associated with pre diabetic state independently of the C-reactive protein. Further studies are needed to support this evidence.


Key words: Prediabetic state; impaired glucose tolerance; impaired fasting glucose; serum sialic acid; protein-bound sialic acid
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## INTRODUCTION

Prediabetes is the term used for individuals with impaired fasting glucose (IFG) and/or impaired glucose tolerance (IGT) and is the main risk factor for the development oftype 2 diabetes mellitus (T2DM) (ADA, 2015, Bergman M, 2013, Eikenberg JD, Davy BM, 2013). Total serum sialic acid (TSA) is a marker of low-grade systemic inflammation as well assignificantly associated with new T2DM (Schmidt MI et al, 1999). However, studies on TSA in those with prediabetes have shown discrepant results. For instance, while sialic acid was not elevated in Melanesians from Fiji with IFG and IGT (Crook MA et al, 2002), an increased TSA in subjects with IGT indicated thatTSA measurement could be valuable as an independent parameter in identifying subjects at higher risk of developing T2DM (Gavella M et al, 2003).
Acylated derivatives of neuraminic acid, called sialic acidare present as terminal components of non-reducing ends of oligosaccharide chains of glycoproteins and glycolipids. Also, sialic acid is localized at the end of many acute phase proteins, asprotein-bound sialic acid (BSA) fraction (Crook MA et al, 2002). However, only small amounts of free sialic acid (FSA) are normally present in serum (Sillanaukee P et al, 1999). Elevated levels of inflammatory markersmay play an important role in prediabetes (Bardini G et al, 2010) consequently, an increased BSA could be expected. Studies on the relationship of both TSA and BSA with metabolic parameters in subjects with prediabetes are scarce (Crook MA et al, 2002, Gavella M et al, 2003, Rajappa M et al, 2013).

The Mexican population has a high prevalence of pre diabetes (Ureña-Bogarín EL et al, 2015), and diabetes (Villalpando S et al, 2010, Aguilar-Salinas CA et al, 2003). In fact, the incidence of T2DMin Mexico is among the highest reported worldwide (González-Villalpando C et al, 2014). Further, diabetes is the number one cause of disability adjusted life years lost in Mexico (Vos T et al, 2013) and fasting and 2h plasma glucose valuesare independent predictors of incident T2DM in Mexican people (Ferrannini E et al, 2009).Therefore, the study of inflammatory markers inMexican subjects with prediabetesis of outmost importance. Toour knowledge, no previous studies have investigated whether there is an increased TSA levelin Mexican adults and/or those with prediabetesor the role that BSA and FSA fractions play.Theaim of the present study was to investigate the sialic acidlevels (TSA, BSA, and FSA) in Mexican adults, and to assess their association with newly diagnosed prediabetes.

## MATERIALS AND METHODS

## Subjects

The participants were selected from 176 adultswho were referred to the outpatient clinic for diabetes of the Dr. Ignacio Chavez Hospital, in Hermosillo, Sonora, Mexico, throughout six months (January to August 2015). As part of their visit to the clinic, all subjects had a medical and biochemical assessment. Participants avoided strenuous exercise and any alcohol intake for two days before the medical and biochemical assessment. From the source population of 176 subjects, 28 were newly diagnosed with prediabetes ( 15 women and 13 men) and agreed to participate in the study. Diagnosis of prediabetes was based on the American Diabetes Association (ADA) criteria (ADA, 2015). Another 31 subjects matched for BMI, sex and age, were selected as controls ( 18 women and 13 men). Subjects with T2DM, hypochromic microcyticanemia, hemoglobinopathies, cancer, hypothyroidism, acute vascular events, hepatic or renal disease, autoimmune diseases or any infection, alcoholism or pregnancywere excluded from the study. In addition, none of them were on anti-hypertensive, lipid-lowering medication, non-steroid antiinflammatories, steroids, oral contraceptive and hormone replacement therapy, pioglitazone, thyroglobulin or fibrates at the time of the study.Thus, 59 subjects with a mean age of $46.15 \pm 12.01$ years met the inclusion criteria for the study and agreed to participate. Written informed consent was obtained from all participants before being included in the study.

This study was approved by the institutional bioethics committee of the Department of Medicine and Health Sciences, University of Sonoraand the bioethics committee of the Dr. Ignacio Chavez Hospital (Dated 13th June 2014). All procedures followed were in accordance with the Declaration of Helsinki 1975, as revised in 2008.

## Analytical methods

The same blood sample used for the biochemical assessment for all 176 subjects was used for this study to measure insulin, PCR and sialic acid in the 59 study participants. Fasting serumlipid profile (total cholesterol, HDLcholesterol, LDL-cholesterol, and triglyceride), total protein, albumin, creatinine, uric acid, alanine aminotransferase (ALT), aspartate aminotransferase (AST) and gamma-glutamyltransferase (GGT) were spectrophotometrically quantified by standardized commercially available diagnostic testsin aHitachi Modular P800 Analyzer (Roche Diagnostics Co., Indiana, USA).LDL-cholesterol was calculated using the Friedewald et al, 1972 formula.

An oral glucose tolerance test (OGTT) was performed after 10-12 h of overnight fasting by using 75 g of glucose as the oral load over a 2 min period, followed by collection of blood samples at baseline and 2 h after the glucose load formeasurement of serum glucose. Results from OGTT were used to define normal glucose tolerance, impaired glucose tolerance or T2DMaccording to ADA criteria (ADA, 2015).
Duplicate $\mathrm{HbA}_{1 \mathrm{c}}$ determinations were performed in a DCA Vantage Analyzer (Siemens DCA Vantage ${ }^{\mathrm{TM}}$ Analyzer, Tarrytown, NY, USA). Fasting serum insulin was determined using a Cal biotech insulin immuno enzymatic assay (Catalog No. IS130D, Spring Valley, CA, USA).Fasting serum glucose was determined using a glucose oxidase assay (Hitachi Modular P800 Analyzer, Roche Diagnostics Co., Indiana, USA). Both measures were combined to calculate the homeostasis model assessment of insulin resistance (HOMA-IR) (Matthews DR, 1985).

Serum C-reactive protein (CRP) was measured in duplicate using a high sensitivity assay by the nephelometric Minineph human CRP assay (hsCRP) (ZK044.L.R, Binding Site Ltd, Birmingham, UK).Duplicate measurements of total serum sialic acid (TSA) and free sialic acid (FSA) were performed using the Quantichrom TM Sialic Acid assay (BioAssay Systems, Hayward, CA). This assay uses an improved Warren method (Warren L, 1959).

## Anthropometric and blood pressure measurements

To obtain body mass index (BMI, $\mathrm{kg} / \mathrm{m}^{2}$ ) weight was measured using an electronic balance (Defender 3000 series, Ohaus, Pine Brook, NJ, U.S.A.) and height was measured with a wall-mounted stadiometer (Holtain Ltd., Dyfed, UK). Waist circumference was measuredusing a flexible tape measure directly on the skin. Participants stood relaxed, with arms folded comfortably across the chest. Measures were made at the end of normal expiration, making sure the tape was positioned perpendicular to the vertical $(\mathrm{Y})$ axis of the body and parallel to the floor.The site was measured twice and the average was used for analysis. Blood pressure was measured using a manual sphygmomanometer, afterthe subject rested for 5 min in a seated position. Systolic and diastolic values were taken as the average of two readings. High blood pressure was defined as a systolic blood pressure at or above 140 mmHg and/or a diastolic blood pressure at or above 90 mmHg , and/or known established hypertension requiring medical treatment.

## Statistical analysis

Either the independent $t$-test or the Mann-Whitney U-test was used to analyze differences between subjects with prediabetes vs. controls for the clinical, biochemical and anthropometric variables, depending whether the variable was normally distributed or not. Data are presented as the arithmetic mean $\pm$ standard deviation (SD), geometric means ( $95 \%$ confidence interval,CI), ormedians (interquartile range, IR), depending on the distribution of the variable. Associations between TSA, BSA and FSA concentrations and prediabetes state were examined using multiple linear regression analysis in separate regression models. Models were constructed using a combination of univariate ( $p<0.2$ ) and stepwise ( $p<0.05$ ) analyses. Interactions of prediabetes state with selected covariates were tested in the models but these were not significant. Collinearity of selected variables was also tested. Model assumptions were tested using a graph of residuals. Statistical analyses were performed using the NCSS 9 statistical software (NCSS 9 Statistical Software 2013, NCSS LCC, Kaysville, UT, USA).

## RESULTS

## Clinical, biochemical and anthropometric characteristics of study groups

Twenty eight subjects (15.9\%) among 176 adultswere identified as newly diagnosed cases of pre diabetes. These subjects shared similar gender and age distributions with the normoglycemic subjects of the control group (Table-1). Significant differences in $\mathrm{HbA}_{1 \mathrm{c}}$, fasting blood glucose and 2-h post-load glucose levels were foundfor the two groups, as expected.Similar body mass index $(\mathrm{BMI})(p=0.076)$ and waist circumference $(p=0.075)$ wereobserved between the two groups. Blood pressure at or above $140 / 80 \mathrm{mmHg}$ was observed in $10.7 \%$ of the subjects with prediabetes and $9.7 \%$ of the control subjects, with no significant difference between the two groups (Table 1).

Insulin concentration was higher ( $p<0.0001$ ) in subjects with prediabetes when compared tocontrols. HOMAIR indexwas also increased in subjects with prediabetes ( $p<0.0001$ ).

Levels of GGT and ALT were within normal ranges (20) in both groups. However, subjects with pre diabetes showed higher concentrations of both enzymesthan controlsubjects ( $p<0.0001$ ). No differences in HDL-cholesterol ( $p=0.223$ ) and triglyceride ( $p=0.676$ ) were observed between groups, although LDL-cholesterol was higher in the control group ( $p=0.007$ ).

Compared to control individuals, subjectswith prediabetes had a lower albumin/globulin ratio ( $p=0.007$ ).Also, higher levels of hs CRP were found in subjectswith prediabetes when compared to controls ( $p=0.035$ ) (Table 1).

Table 1. Clinical, biochemical and anthropometric variables of the subjects with prediabetes and control subjects at baseline.

| Variable | Subjects with prediabetes ( $\mathrm{n}=28$ ) | Control subjects $(\mathrm{n}=31)$ | $P$ value |
| :---: | :---: | :---: | :---: |
| Gender (\%Men) | 44.8 | 43.3 | 0.799 |
| Age (years) | $46.4 \pm 11.3$ | $45.9 \pm 12.8$ | 0.854 |
| Altered blood pressure (\%) | 10.7 | 9.7 | 0.1500 |
| HbA1c, \% (mmol/mol) | $6.0 \pm 0.3(42.6 \pm 3.3)$ | $5.2 \pm 0.3(33.5 \pm 3.5)$ | $<0.0001$ |
| Fasting glucose (mg/dL) | $99.8 \pm 11.1$ | $84.7 \pm 5.6$ | <0.0001 |
| 2-h glucose (mg/dL) | $140.5 \pm 30.1$ | $112.1 \pm 22.6$ | <0.0001 |
| Fasting insulin ( $\mu \mathrm{IU} / \mathrm{mL})^{\text {a }}$ | 5.0 (4.1, 6.3) | 3.17 (2.7, 3.8) | <0.0001 |
| HOMA-IR ${ }^{\text {a }}$ | 5.0 (4.1, 6.3) | 3.17 (2.7, 3.8) | <0.0001 |
| Total sialic acid (mg/dL) | $76.8 \pm 10.3$ | $61.9 \pm 9.0$ | <0.0001 |
| Bound sialic acid (mg/dL) | $76.3 \pm 10.2$ | $61.4 \pm 9.0$ | $<0.0001$ |
| Free sialic acid (mg/dL) ${ }^{\text {b }}$ | $0.5(0.1,0.5)$ | $0.5(0.2,0.7)$ | 0.443 |
| Total cholesterol (mg/dL) ${ }^{\text {b }}$ | 187.0 (47.0, 141.0) | 201.0 (24.3, 162.0) | 0.096 |
| Triglyceride (mg/dL) | 142.9 (57.6) | 137.5 (37.7) | 0.676 |
| HDL-cholesterol (mg/dL) | $47.3 \pm 9.4$ | $50.5 \pm 10.8$ | 0.223 |
| LDL-cholesterol (mg/dL) | $106.8 \pm 34.7$ | $131.2 \pm 33.0$ | 0.007 |
| Total proteins (g/dL) | $7.0 \pm 0.4$ | $6.9 \pm 0.5$ | 0.448 |
| Globulins (g/dL) | $3.1 \pm 0.5$ | $2.8 \pm 0.8$ | 0.039 |
| Albumin/Globulin ratio (g/dL) ${ }^{\text {a }}$ | 1.2 (1.1, 1.3) | $1.5(1.3,1.8)$ | 0.007 |
| hsCRP (mg/L) ${ }^{\text {b }}$ | 1.6 (3.2, 10.1) | 1.0 (1.1, 7.1) | 0.035 |
| Gamma-glutamyltransferase (IU/L) ${ }^{\text {a }}$ | 20.6 (16.3, 26.1) | 9.4 (7.8, 11.2) | <0.0001 |
| Aspartateaminotransferase (IU/dL) ${ }^{\text {a }}$ | 15.3 (13.7, 17.1) | 13.5 (11.9, 15.4) | 0.431 |
| Alanineaminotransferase (IU/dL) ${ }^{\text {b }}$ | 18.5 (12.9, 41.0) | 12.6 (5.5, 57.6) | <0.0001 |
| Height (cm) ${ }^{\text {b }}$ | 161.1 (13.0, 32.2) | 164.4 (18.2, 180.5) | 0.933 |
| Weight (kg) | $85.8 \pm 17.7$ | $80.3 \pm 13.3$ | 0.176 |
| Body mass index (kg/m²) | $31.7 \pm 5.2$ | $29.57 \pm 3.8$ | 0.076 |
| Waist circumference (cm) | $107.4 \pm 12.6$ | $101.7 \pm 11.4$ | 0.075 |

A t test was done for independent samples. Non-normal distribution data were tested with the Mann-Whitney U-test.
${ }^{\text {a }}$ Geometric means ( $95 \%$ CI)
${ }^{\mathrm{b}}$ Medians (IR)
${ }^{\text {c }}$ All other data are presented as mean $\pm \mathrm{SD}$

* Altered blood pressure $>130 / 90 \mathrm{mmHg}$
hs CRP, high sensitive C-reactive protein


## Quantification of the different forms of sialic acid

Mean TSA and BSAlevelswere higher in subjects with pre diabetes than control subjects ( $p<0.0001$ and $p<0.0001$, respectively). However, no significant differences between groups were observed for serum FSA level ( $p=0.443$ ) (Table 1).

## Univariate analysis of serum sialic acid forms

Univariate regression analyses were performed with TSA, BSA or FSA as the dependent variables. Results showed positive association of serum TSA with fasting insulin ( $p=0.0031$ ), HOMA-IR ( $p=0.0018$ ), GGT ( $p<0.0001$ ), hs CRP ( $p=0.0008$ ), total proteins ( $p=0.0058$ ), globulins ( $p=0.0070$ ), BMI ( $p=0.0019$ ), and waist circumference ( $p=0.0464$ ); and a negative association withthe albumin/globulin ratio ( $p=0.0143$ )(Table 2).Likewise,results showed a positive association of BSAwith fasting insulin ( $p=0.0033$ ), HOMA-IR ( $p=0.0019$ ), GGT ( $p<0.0001$ ), hs CRP ( $p=0.0009$ ), total proteins ( $p=0.0058$ ), globulins ( $p=0.0067$ ), BMI ( $p=0.0019$ ), and waist circumference ( $p=0.0473$ ); and a negative association with the albumin/globulin ratio ( $p=0.0137$ ) (Table 2). FSA was positively associated only with triglyceride ( $p=0.0088$ ), LDL cholesterol ( $p=0.0118$ ), tri/HDL ratio ( $p=0.0230$ ) and creatinine ( $p=0.0098$ ) (Table 3).

## Multiple linear regression analysis of serum sialic acid and associated variables

Multiple regression models confirmed the association of TSA and BSA with prediabetes state (TSA: $\beta=11.9$, $p<0.0001$; BSA: $\beta=12.0, p<0.0001$ ) (Table 4). Subjects with prediabetes showedhigher levels of TSA and BSA than those with normal glucose levels after adjusting for hs CRP, total proteins and AST.

Table 2.Univariate relationships between total sialic acid and bound sialic acid with clinical, biochemical and anthropometric characteristics in the whole group ( $\mathrm{n}=59$ ).

| Variable | Total Sialic Acid |  |  | Bound Sialic Acid |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\beta$ | Standard <br> Error | $p$ | $\beta$ | Standard <br> Error | $p$ |
| Prediabetes (yes) | 14.9 | 2.5 | $<0.0001$ | 14.9 | 2.5 | $<0.0001$ |
| Gender (men) | -4.1 | 3.2 | 0.1966 | -4.2 | 3.1 | 0.1904 |
| Altered blood pressure (\%) | 7.2 | 3.7 | 0.0581 | 7.2 | 3.7 | 0.0585 |
| HbA $_{1 \mathrm{l}}$ \% (mmol/L) | 1.5 | 0.2 | $<0.0001$ | 1.5 | 0.2 | $<0.0001$ |
| Fasting glucose (mg/dL) | 0.5 | 0.1 | 0.0003 | 0.5 | 0.1 | 0.0003 |
| 2-h glucose (mg/dL) | 0.1 | 0.1 | 0.0393 | 0.1 | 0.1 | 0.0403 |
| Fasting insulin ( $\mu \mathrm{IU} / \mathrm{mL})$ | 1.6 | 0.5 | 0.0031 | 1.6 | 0.5 | 0.0033 |
| HOMA-IR | 6.2 | 1.9 | 0.0018 | 6.2 | 2.0 | 0.0019 |
| HDL-cholesterol (mg/dL) | -0.3 | 0.6 | 0.0829 | -0.3 | 0.2 | 0.0850 |
| Total proteins (g/dL) | 9.5 | 3.3 | 0.0058 | 9.5 | 3.3 | 0.0058 |
| Globulins (g/dL) | 6.4 | 2.3 | 0.0070 | 6.5 | 2.3 | 0.0067 |
| Albumin/Globulin ratio (g/dL) | -7.1 | 2.8 | 0.0143 | -7.2 | 2.8 | 0.0137 |
| hsCRP (mg/L) | 2.2 | 0.6 | 0.0008 | 2.2 | 0.6 | 0.0009 |
| Gamma-glutamyltransferase <br> (IU/L) | 0.5 | 0.1 | $<0.0001$ | 0.5 | 0.1 | $<0.0001$ |
| Aspartateaminotransferase <br> (IU/dL) | 0.5 | 0.2 | 0.0714 | 0.5 | 0.2 | 0.0729 |
| Alanineaminotransferase <br> (IU/dL) | 0.4 | 0.1 | 0.0121 | 0.4 | 0.1 | 0.0121 |
| Weight (kg) | 0.2 | 0.1 | 0.1199 | 0.1 | 0.1 | 0.1236 |
| Body mass index (kg/m ${ }^{2}$ ) | 1.1 | 0.3 | 0.0019 | 1.1 | 0.3 | 0.0019 |
| Waist circumference (cm) | 0.3 | 0.1 | 0.0464 | 0.3 | 0.1 | 0.0473 |

Table 3. Univariate relationship between free sialic acid and clinical, biochemical and anthropometric characteristics of the study groups ( $\mathrm{n}=59$ ).

|  | Free Sialic Acid |  |  |
| :---: | :---: | :---: | :---: |
| Variable | $\beta$ | Standard Error | p |
| Group (prediabetic group yes) | -0.0494 | 0.0358 | 0.1738 |
| Gender (men) | 0.0613 | 0.0357 | 0.0915 |
| Cholesterol (mg/dL) | 0.0008 | 0.0005 | 0.1109 |
| Triglyceride $(\mathrm{mg} / \mathrm{dL})$ | 0.001 | 0.0004 | 0.0088 |
| LDL cholesterol $(\mathrm{mg} / \mathrm{dL})$ | 0.0013 | 0.0005 | 0.0118 |
| Triglyceride/HDL ratio $(\mathrm{mg} / \mathrm{dL})$ | 0.0317 | 0.0136 | 0.0230 |
| Creatinine $(\mathrm{mmol} / \mathrm{L})$ | 0.3175 | 0.1188 | 0.0098 |

Table 4.Multiple regression models for total sialic acid, bound sialic acid and free sialic acid inthe study groups ( $\mathrm{n}=59$ ).

|  | Total Sialic Acid |  | Bound Sialic Acid |  | Free Sialic Acid |  |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\beta$ | $p$ | $\beta$ | $p$ | $\beta$ | $p$ |
| Prediabetes (Yes) | 11.9 | $<0.0001$ | 12.0 | $<0.0001$ | -0.04 | 0.1591 |
| C reactive protein (mg/L) | 1.1 | 0.0267 | 1.1 | 0.0275 | - | - |
| Total proteins $(\mathrm{g} / \mathrm{dL})$ | 8.1 | 0.0017 | 8.1 | 0.0017 | - | - |
| Aspartate aminotransferase (IU/dL) | 0.4 | 0.0230 | 0.4 | 0.0238 | - | - |
| Cholesterol (mg/dL) | - | - | - | - | -0.002 | 0.0028 |
| Triglycerides (mg/dL) | - | - | - | - | 0.001 | 0.0001 |
| LDL-Choleterol (mg/dL) | - | - | - | - | 0.0031 | 0.0012 |
| Creatinine (mmol/L) | - | - | - | - | 0.3 | 0.0007 |

## DISCUSSION

In this study, prediabetes was detected in $15.9 \%$ of 176 adults who participated as the source group. Similar results were reported from a cross-sectional study on Mexican adult population (Ureña-Bogarín EL et al, 2015).

Some studies have shown increased TSA in subjects with IGT (Gavella M et al, 2003). However, studies on the relationship between TSA fractions and prediabetes are scarce (Rajappa M et al, 2013). Data from the present study showed significantly increased serum TSA and BSA concentrations in subjects with prediabetes, compared to control subjects. The lack of differences in FSA concentrations between groups suggests this fraction of sialic acid does not contribute to the increase of TSA.In addition, a direct association between BSA and prediabetes was also demonstrated.

In this context, Rajappa et al. 2013 showed that BSA is elevated in non-diabetic obese subjects, and concluded that BSA levels are associated with insulin resistance and obesity. An association between BSA and prediabetes was not found (Rajappa M et al, 2013). This is important because when analyzing data using the ADA, 2015 criteria in the aforementioned study, it wasobserved that several individuals had altered fasting plasma glucose.

As elevation in serum sialic acid occurs in low-grade systemic inflammation, the increment of acute phase proteins would be associated with BSAlevelsin subjects with prediabetes. In human beings the most derivative (90\%) sialic acid is linked to globulins (Carter A et al, 1962) in the form of BSA. Accordingly, in our study asignificant rise inCRPin subjects with prediabetes was observed when compared to control subjects. These results are similar to those reported by (Sabanayagam et al.2011), who demonstrated that CRP levels are associated with prediabetes.Also, in the presentstudyonlythe subjects with prediabetes showed CRP levels above the low risk referent of $1 \mathrm{mg} / \mathrm{L}$ (Pearson TA et al, 2003), which suggest they have a higher risk for chronic subclinical inflammation than control subjects.

Differences in CRP concentrations between subjects with prediabetes and controlsubjects could be associated to the increased sialic acid concentrations, due to the fact that CRP is part of the acute-phase response. Since CRP is a non-glycosylated protein (Schultz DR, Arnold PI, 1990) and sialic acid is bound to most acute-phase glycoproteins (Sillanaukee P et al, 1999), differences in concentrations of CRP in the current study could partiallyexplain the rise of BSA levels in subjectswith prediabetes.

In the present work, the association between sialic acid concentration andprediabetes proved to be independent ofCRP levels.It has been reported that, compared to CRP, TSA is a more robust inflammatory marker and a better predictor of metabolic-associated inflammation (Browning et al. 2004). An explanation could be that CRP is subject to greater individual variation. Furthermore, serum TS Amay reflect the global acute-phase responseproduced by the increased synthesis of glycoproteins (Pepys MB, 2003) whereas CRP represents the response of only one protein that is not sialylated (Pepys MB, 2003).

At the same time,subjectswith prediabetes had higher HOMA-IR than controls.However, HOMA-IR was not independently associated with sialic acidlevels.In contrast, (Browning et al. 2004) demonstrated that TSA was directly associated with insulin resistance and BMI but also with dislypidemia and the hypertensive state of patients with metabolic syndrome. This discrepancycould be explained by the fact that subjects and controls in our study had similar BMI, waist circumference, and blood pressure.

In conclusion, the present studyshowed increased TSA and BSA levels in Mexican adults with newly diagnosed prediabetes. Increased TSA relied on the increment of BSA, and both TSA and BSA were associated with prediabetes independently of the CRPlevel. Further research is required to elucidate the reason for the increased BSA levels in subjects with prediabetes. Since BSA may increase as a result of an increased acute-phase response, additional studies are required to look for alterations inother hepatic glycoproteins, besides CRP. Overall, the results and the evidence created from this study may be considered reliable from the point of view of its design. However it has the limitation of the relatively low number of participants. Therefore, further studies are needed to support this evidence.

## Conflicts of interest

No potential conflicts of interest relevant to this article were reported.
M. C. Candia-Plata designed the study, oversaw the conduct of the study, conducted the clinical and biochemical assessment, wrote the paper, and is the guarantor. A.L. Mata-Pineda and H. Valdez-Salazarrecruited subjects and conducted clinical assessment. Also A.L. Mata-Pineda performed the anthropometric and biochemical measurements. M.L. González-Félix assisted the study and writing the manuscript. J. Esparza-Romero, G. Alvarez and E. Valencia assisted the design of the study and carried out the statistical analysis and contributed to writing the paper. The authors thank the patients who volunteered to participate in this study.

## REFERENCES

Aguilar-Salinas CA, Velazquez MO, Gómez-Pérez FJ, Gonzalez CA, Lara EA, Molina CA, (2003). Characteristics of patients with type 2 diabetes in Mexico results from a large population-based nationwide survey. Diabetes Care, 26 (7): 2021-2026.
American Diabetes Association. (2015). Classification and Diagnosis of Diabetes. Diabetes Care. 38 (Suppl 1): S816.

Bardini G, Dicembrini I, Cresci B, Rotella CM. (2010). Inflammation markers and metabolic characteristics of subjects with 1-h plasma glucose levels. Diabetes Care. 33 (2): 411-413.
Bergman M. (2013). Pathophysiology of prediabetes and treatment implications for the prevention of type 2 diabetes mellitus. Endocrine. 43 (3): 504-513.
Browning LM, Jebb SA, Mishra GD, Cooke JH, O'connell MA, Crook MA. (2004). Elevated sialic acid, but not CRP, predicts features of the metabolic syndrome independently of BMI in women. Int J Obesity. 28: 10041010.

Carter A, Martin NH. (1962). Serum sialic acid levels in health and disease. J ClinPathol. 15:69-72.
Crook MA, Goldsmith L, Ameerally P, Lumb P, Singh N, Miell J. (2002). Serum sialic acid, a possible cardiovascular risk factor is not increased in Fijian Melanesians with impaired glucose tolerance or impaired fasting glucose. Ann Clin Biochem. 39 (6): 606-608.
Crook M. (1993).The determination of plasma or serum sialic acid. Clin Biochem. 1: 31-8.
Eikenberg JD, Davy BM. (2013). Prediabetes: A prevalent and treatable, but often unrecognized clinical condition. J Acad Nutr Diet. 113 (2): 213-218.
Ferrannini E, Massari M, Nannipieri M, Natali A, Ridaura RL, Gonzales-Villalpando C. (2009). Plasma glucose levels as predictors of diabetes: the Mexico City diabetes study. Diabetologia. 52 (5): 818-24.
Friedewald WT, Levy RI, Fredrickson DS. (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem. 18 (6): 499-502.
Gavella M, Lipovac V, Car A, Vučić M, Sokolić L, Rakoš R. (2003). Serum sialic acid in subjects with impaired glucose tolerance and in newly diagnosed type 2 diabetic patients. Acta Diabetol. 40: 95-100.
González-Villalpando C, Dávila-Cervantes CA, Zamora-Macorra M, Trejo-Valdivia B, González-Villalpando ME. (2014). Incidence of type 2 diabetes in Mexico: Results of the Mexico City Diabetes Study after 18 years of follow-up. Salud Publica Mex. 56 (1): 11-17.
Lee DH, Silventoinen K, Jacobs Jr DR, Jousilahti P, Tuomileto J. (2004). $\gamma$-Glutamyltransferase, obesity, and the risk of type 2diabetes: observational cohort study among 20,158 middle-aged men and women. J Clin Endocrinol Metab. 89 (11): 5410-5414.
Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. (1985). Homeostasis model assessment: insulin resistance and $\beta$-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia. 28 (7):412-419.

Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon III RO, Criqui M. (2003). Markers of inflammation and cardiovascular disease application to clinical and public health practice: a statement for healthcare professionals from the centers for disease control and prevention and the American Heart Association. Circulation. 107 (3):499-511.
Pepys MB, (2003). Hirschfield GM.C-reactive protein: a critical update. J Clin Invest. 111 (12): 1805-1812.
Rajappa M, Ikkruthi S, Nandeesha H, Satheesh S, Sundar I, Ananthanarayanan P H. (2013). Relationship of raised serum total and protein bound sialic acid levels with hyperinsulinemia and indices of insulin sensitivity and insulin resistance in non-diabetic normotensive obese subjects. Diabetes Metab Syndr. 7 (1): 17-19.
Sabanayagam C, Shankar A, Lim SC, Lee J, Tai ES, Wong TY. (2011). Serum C-reactive protein level and prediabetes in two Asian populations. Diabetologia. 54 (4): 767-775.
Schmidt MI, Duncan BB, Sharrett AR, Lindberg G, Savage PJ, Offenbacher S, (1999). Markers of inflammation and prediction of diabetes mellitus in adults (Atherosclerosis Risk in Communities study): a cohort study.The Lancet. 353 (9165): 1649-1652.
Schultz DR, Arnold PI. (1990). Properties of four acute phase proteins: C-reactive protein, serum amyloid A protein, alpha 1-acid glycoprotein, and fibrinogen. Seminars in Arthritis and Rheumatism. 3:.129-147.
Sillanaukee P, Ponnio M, Jääskeläinen IP. (1999). Occurrence of sialic acids in healthy humans and different disorders. Eur J Clin Invest. 29 (5): 413-425.
Ureña-Bogarín EL, Martínez-Ramírez HR, Torres-Sánchez JR, Hernández-Herrera A, Cortés-Sanabria L, CuetoManzano AM. (2015). Prevalence of pre-diabetes in young Mexican adults in primary health care. Fam Pract. 32 (2): 159-164.
Villalpando S, De la Cruz V, Rojas R, Shamah-Levy T, Ávila MA, Gaona B, (2010). Prevalence and distribution of type 2 diabetes mellitus in Mexican adult population: a probabilistic survey. Salud Publica Mex. 52: S19S26.
Vos T, Barber RM, Bell B, Bertozzi-Villa A, Biryukov S, Bolliger I. (2013). Global, regional, and national incidence, prevalence, and years lived with disability for 301 acute and chronic diseases and injuries in 188 countries, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. The Lancet. 386 (9995): 743-800.

Warren L. (1959). The thiobarbituric acid assay of sialic acids. J Biol Chem. 234 (8): 1971-1975.
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