

## Review Article

---

# Colitis Nucleomigrans: A Proposal for The Third Type of Microscopic Colitis

Mitsuhiro Tachibana, MD<sup>1\*</sup>, Yutaka Tsutsumi, MD<sup>1,2</sup>

<sup>1</sup>Department of Diagnostic Pathology, Shimada Municipal Hospital, Shimada, Shizuoka, Japan

<sup>2</sup>Diagnostic Pathology Clinic, Pathos Tsutsumi, Inazawa, Aichi, Japan

**\*Corresponding Author:** Dr. Mitsuhiro Tachibana, Department of Diagnostic Pathology, Shimada Municipal Hospital, 1200-5 Noda, Shimada, Shizuoka 427-8502, Japan, Tel: +81-547-35-2111 (ext. 2273); Fax: +81-547-35-2182; ORCID ID: <https://orcid.org/0000-0002-1041-0391>

**Received:** 12 March 2021; **Accepted:** 22 March 2021; **Published:** 31 March 2021

## Abstract

Recently, we proposed the third entity of microscopic colitis (MC), termed colitis nucleomigrans (CN). The present review describes clinicopathological features of CN. CN shares clinical and endoscopic features of MC with collagenous colitis and lymphocytic colitis. We analyzed endoscopic biopsy specimens of nonspecific colitis clinically manifesting chronic watery diarrhea or inflammatory bowel disease (IBD)-like symptoms, but with minor endoscopic abnormality. The histopathological criteria of CN are as follows: a) chained nuclear migration to the middle part of the surface-lining columnar epithelia, b) apoptotic nuclear debris scattered below the nuclei, and c) mild to moderate chronic inflammation in the lamina propria. Thirty-three patients (M:F=20:13, median 63 years; range 17-88) fulfilled the above criteria. Seven cases accompanied MC-like clinical/endoscopic features. Mucosal reddening with or without erosions/apthae was endoscopically observed in the remaining 26 cases with IBD-like clinical features: occult/gross hematochezia (n=19), abdominal pain (n=2) and mucin secretion (n=2). Apoptotic debris immunoreactive for cleaved caspase-3 appeared more frequently in IBD-like CN than in MC-like CN. CD8-positive intraepithelial lymphocytes were comparable in both types. Proton pump inhibitors (PPIs) were administered in five (71%) CN cases with MC-like features, and the diarrhea improved after cessation of PPIs in three. In IBD-like CN cases, eight (31%) received PPIs. Altered apoptotic processes in the colorectal surface-lining epithelia, predominantly with a debris pattern of

apoptosis, may be involved in the pathogenesis. Mechanisms of nuclear migration to the unusual position and the impairment of nuclear anchoring to the basal situation in the surface-lining epithelia remain to be established.

**Keywords:** Apoptosis; Colitis nucleomigrans; Colorectal diseases; Watery diarrhea; Intraepithelial lymphocytes; Microscopic colitis

**Abbreviations:** CN- Colitis nucleomigrans; CC- Collagenous colitis; IBD- Inflammatory bowel disease; LC- Lymphocytic colitis; MC- Microscopic colitis; NSAIDs- Non-steroid anti-inflammatory drugs; PPI- Proton pump inhibitor

## **1. Introduction**

Chronic colitis can be divided into the following categories; i.e. inflammatory bowel disease (IBD), treatment-related colitis (pseudomembranous colitis, chemical colitis, chemotherapy-induced colitis, and radiation colitis), infectious colitis, ischemic colitis, and microscopic colitis (MC) [1]. The appropriate microscopic diagnosis of colitis is made by diagnostic pathologists who give routine pathology services. MC, predominantly seen in middle-aged women, is characterized by chronic non-bloody watery diarrhea and an endoscopically normal but microscopically altered specific appearance of the colonic mucosa [2,3]. Other common symptoms include nocturnal diarrhea, abdominal pain and weight loss [2, 3].

The category of MC has encompassed two distinct entities such as collagenous colitis (CC) and lymphocytic colitis (LC). They share clinical manifestations and are occasionally associated with autoimmune diseases [3-5]. Histologically, CC is featured by the deposition of subepithelial collagen fibers “collagen bands” in the uppermost part of the lamina propria mucosae, while LC characteristically reveals marked increase of CD8-positive intraepithelial lymphocytes (IELs). Overlapped lesions between CC and LC have been reported [3-5].

In daily diagnostic pathology practice, we often encounter biopsy specimens that display microscopic features of chronic nonspecific colitis without collagen bands or increased IELs. Recently, we proposed the third type of MC, colitis nucleomigrans (CN) [6,7]. Histologically, the nuclei of surface-lined colonic columnar cells migrate in chains to the middle part of the cells, and fragmented debris of apoptotic nuclei is scattered beneath the migrated nuclei. A total of 33 cases were analyzed clinicopathologically. The patients revealed chronic non-bloody watery diarrhea but with normal endoscopic appearance (MC-like symptoms) (n=7) or microscopic/gross bloody stools with endoscopic mucosal redness, abdominal pain and/or mucin secretion (IBD-like symptoms) (n=26).

## **2. Clinical and endoscopic features of CN**

Thirty-three patients satisfied the microscopic criteria of CN [6]: a) chained nuclear migration to the middle part of the surface-lining columnar epithelium, b) apoptotic nuclear debris scattered below the migrated nuclei, and c) mild to moderate infiltration of lymphocytes and plasma cells in the lamina propria. The frequency of CN among colorectal biopsy specimens was 33/1287 (2.6%). Twenty men and 13 women were enrolled, and the age ranged from 17 to 88 years with the mean 58.4 and the median 63. Patients manifested chronic non-bloody watery diarrhea (MC-like symptoms) with normal endoscopic appearance (n=7; Figure 1) or occult (n=9)/gross (n=10) haematochezia, abdominal pain (n=2) or mucin secretion (n=2) (IBD-like symptoms), endoscopically showing mucosal reddening with or without focal erosions/apthae (n=26; Figure 2). One patient in the MC-like group exceptionally accompanied fecal occult blood, while normal-looking endoscopic findings were recorded in three patients in the IBD-like group.

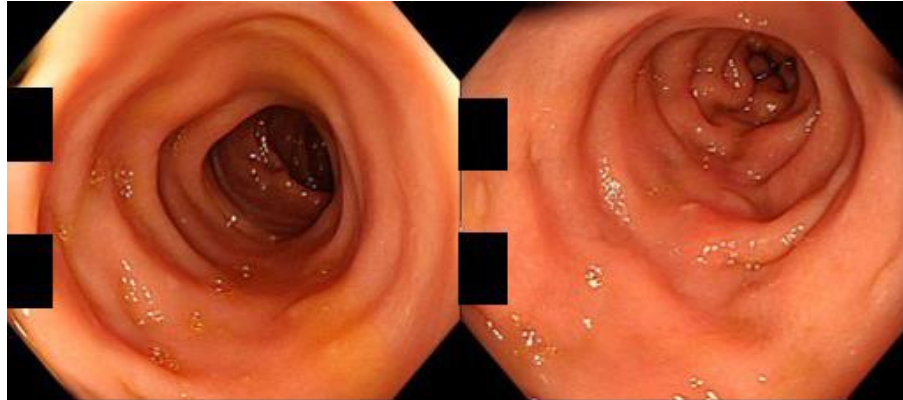
PPIs were regularly administered in 13 patients; five (71%) with MC-like features and eight (31%) with IBD-like features. In three in the MC-like group, diarrhea improved following the discontinuation of medication. H<sub>2</sub>-blockers and non-steroidal anti-inflammatory drugs (NSAIDs) were used in two patients each in the IBD-like group. Hypertension was documented in eight patients. Two patients were suspected of Behçet disease, but without intestinal ulceration. One patient suffered from IgA nephropathy, and one patient suffered Sjögren syndrome. Four patients underwent chemotherapy against plasma cell myeloma, nodal diffuse large B-cell lymphoma, splenic diffuse large B-cell lymphoma, or colonic mucinous adenocarcinoma. The periods between the completion of chemotherapy and the diagnosis of CN were 0, 5, 6 and 53 months, respectively.

Clinical and endoscopic features of CN with MC-like features (n=7) and CN with IBD-like features (n=26) are summarized in Table 1.

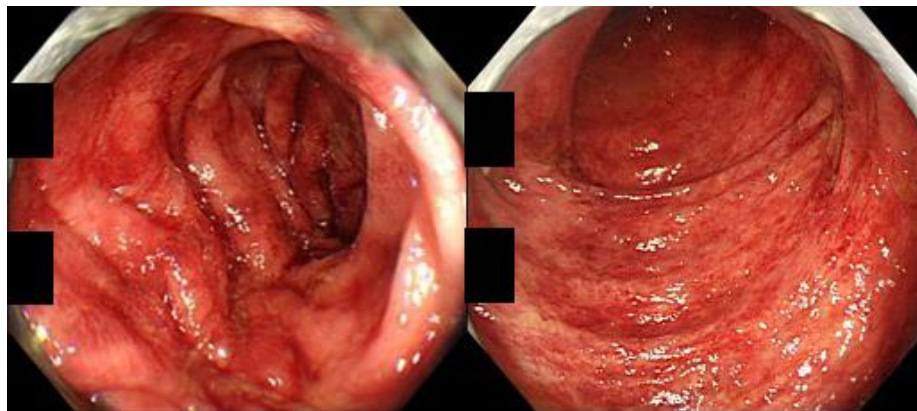
<b>i) Cases with microscopic colitis-like features (n=7)</b>						
Age (yrs)/Gender	Watery diarrhoea	IBD-like features	Endoscopic findings	C-caspase 3(/100 epithelial cells)	CD8 <sup>+</sup> IELs (/100 epithelial cells)	Medication
26–80	3+: 7	FOB: 1, none: 6	normal: 6	1.4–22.6	2.3–17.4	Esomeprazole: 2*
average: 58			mucosal edema: 1	average: 10.3	average: 7.0	Lansoprazole: 2
median: 65				median: 9.9	median: 3.6	Rabeprazole: 1*
M:F=4:3						
<b>ii) Cases with inflammatory bowel disease-like features (n=26)</b>						
Age (yrs)/Gender	Watery diarrhea	IBD-like features	Endoscopic findings	C-caspase 3(/100 epithelial cells)	CD8 <sup>+</sup> IELs (/100 epithelial cells)	Medication
17–88	0:23	FOB: 9	normal: 3	1.8–64.8	1.6–17.2	Rabeprazole: 4
average: 58.5	1+: 1	melena: 10	reddening: 16	average: 24.3	average: 7.6	Lansoprazole: 2
median: 61	2+: 1	abdominal pain: 1	erosion: 3	median: 23.6	median: 7.5	Esomeprazole, 2
M:F=16:10	3+: 1	abdominal pain & mucin secretion: 1	FIV & MR: 1			Vonoprazan: 1 NSAIDs: 2
		mucin secretion: 1	ischemia-like change: 2			H <sub>2</sub> blocker: 2
		weight loss: 2	reddening			NSAIDs
		detailed check for colon cancer: 1	erosion & aphtha			
		detailed check for Behçet disease: 1				

Abbreviations: M, male; F, female; yrs, years; watery diarrhea score: 0, negative; 1+, mild; 2+, moderate; 3+, severe IBD, inflammatory bowel disease; C-caspase 3, cleaved-caspase 3; IELs, intraepithelial lymphocytes; FOB, fecal occult blood; NSAIDs, non-steroidal anti-inflammatory drugs; FIV, focal indistinct vascular pattern; MR, mucosal roughening; \* diarrhoea improved after cessation of PPIs.

**Table 1:** Summary of cases of colitis nucleomigrans.



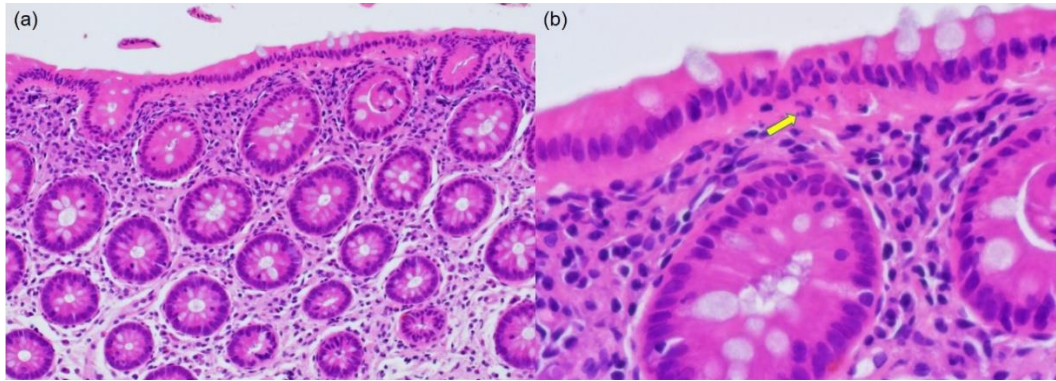
**Figure 1:** Colonoscopic findings in a patient of CN with MC-like symptoms (80-year-old man; left: transverse colon, right: sigmoid colon). The colorectal mucosa appears normal without mucosal edema, reddening, erosion or ulceration.



**Figure 2:** Colonoscopic findings in a patient of CN with IBD-like symptoms (82-year-old man; left: sigmoid colon, right: rectum). The colonic mucosa displays multiple patchy reddening.

### 3. Histopathological features of CN

The total number of biopsy pieces with microscopic features of CN sampled from 33 cases reached 65. The microscopic features of CN were observed in any part of the large intestine (cecum 3, ascending colon 7, transverse colon 10, descending colon 8, sigmoid colon 21 and rectum 16). The chained nuclear migration was never experienced in the crypt epithelial cells. Collagen bands and increased IELs (>20 IELs among 100 epithelial cells) failed to be noted in the specimens. Representative microscopic appearance of CN is demonstrated in Figure 3.

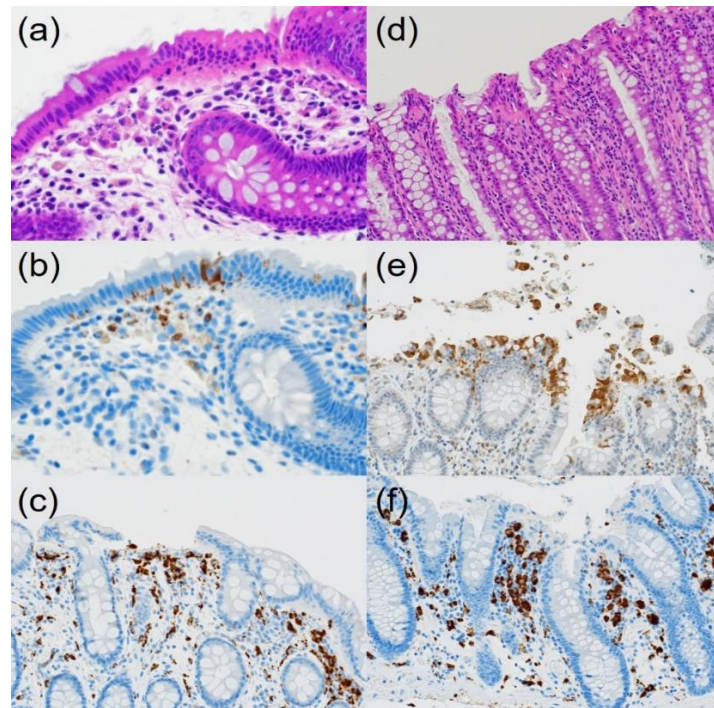


**Figure 3:** Microscopic findings of CN with IBD-like symptoms (54-year-old man; HE). (a) The nuclei of the surface-lining columnar cells are migrated in chain to the middle part of the cells. Eosinophilic cytoplasm is evident under the chained nuclei. The colonic crypts are arranged in parallel without features of cryptitis or nuclear migration. The lamina propria mucosae reveals moderate infiltration of lymphocytes and plasma cells. (b) A high-powered view shows the migrated chained nuclei in the surface-lining columnar epithelial cells. Under the nuclei, apoptotic bodies (clustered fragments of nuclear debris) are seen (arrow). Subepithelial collagen bands and increased IELs are not recognized.

In the period of the present study, we experienced only five cases of CC and no case of LC. The colorectal mucosa in a remission state of UC (n=20), as well as normal colorectal mucosa sampled from surgical specimens of colorectal cancer (n=5), were also evaluated. The CN-type microscopic features were not seen in CC and control normal colorectal mucosa, but observed in twelve (60%) lesions in UC in a remission state. IELs were immunoreactive for both CD3 and CD8. IELs were scattered among the CN lesion (but without significant increase). The average numbers of CD8-positive IELs were 7.0 per 100 surface epithelial cells (median: 3.6, range: 2.3-17.4) in CN with MC-like features, and 7.9 (median: 7.9, range: 1.6-17.2) in CN with IBD-like features. In the control group, the mean numbers of IELs were 18.1 (median: 7.2, range: 4.0-64.9) in CC, 7.6 (median: 6.0, range: 0-18.3) in UC in a remission state, and 4.4 (median: 4.2, range: 1.3-7.1) in surgically resected normal colorectal mucosa. Apoptotic bodies with a ‘debris pattern’ (clustered fragments of nuclear debris) beneath the migrated nuclei in CN were clearly immunoreactive for cleaved caspase-3 (Figure 4a and 4b). In the MC-like CN lesions, cleaved caspase-3-positive apoptotic bodies of debris type counted 10.3 per 100 surface-lining epithelial cells on average (median: 9.9, range: 1.4-22.6), while the average number in the IBD-like CN lesions was higher: 28.2 (median: 20.5, range: 1.8-64.8).

In the surgically resected normal colorectal mucosa, cleaved caspase-3-immunoreactivity was seen mainly as non-fragmented apoptotic nuclei along the luminal space on the surface-lining cells, representing an ‘extrusion pattern’ of apoptosis (Figure 4c and 4d). Apoptosis of debris type was infrequently observed in normal colorectal mucosa. Apoptosis of extrusion type was occasionally seen also in the CN lesions. The number of cleaved caspase 3-

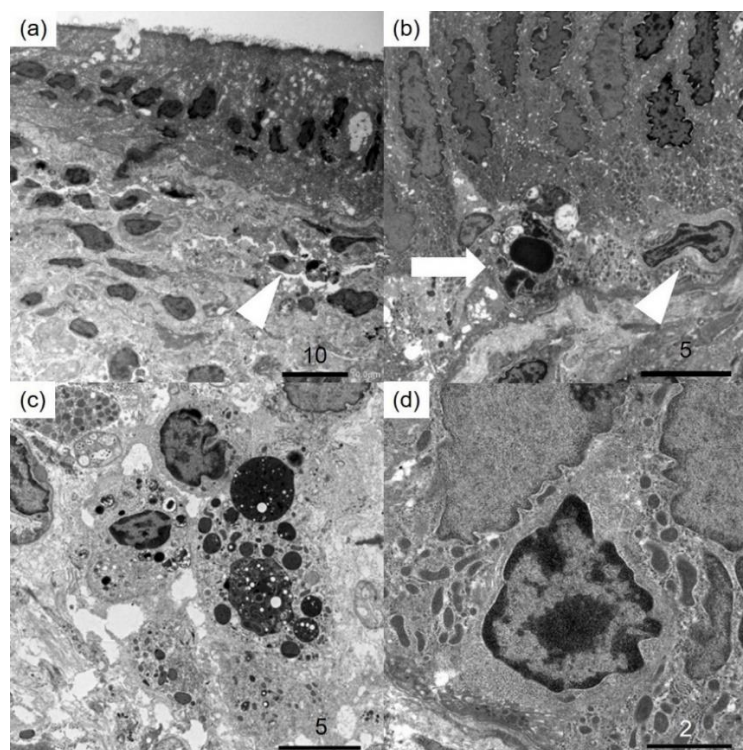
immunoreactive apoptotic bodies of extrusion type in the normal colorectal mucosa ranged from 1.0 to 5.4 (mean 3.4, median: 3.5) per 100 epithelial cells. The apoptotic bodies in CC (n=5) were predominantly categorized in the debris type and their number ranged from 6.1 to 23.4 (mean: 14.3, median: 13.5). In the lesions in UC in a remission state, epithelial apoptosis was identified in mixed debris and extrusion patterns: The number of apoptosis of the debris type ranged from 6.2 to 63.2 (mean: 29.1, median 28.2). CD68-positive macrophages located in the upper part of the lamina propria phagocytized cleaved caspase 3-immunoreactive apoptotic bodies (fragmented nuclear debris) in the cytoplasm. Such a pattern was observed in any type of the colorectal lesions and in normal colorectal mucosa, but their number was increased in the lesion of CN (Figure 4e and 4f). The CD68-positive macrophages in CN showed the plump and amoeboid cytoplasm, while those in the normal colorectal mucosa tended to be round and smaller in size. The number of apoptotic debris-phagocytizing CD68-positive macrophages in CN with IBD-like features was comparable with that in CN with MC-like features.



**Figure 4:** Comparative immunohistochemical features of CN (a-c, 40-year-old woman) and control normal colon (d-f, 82-year-old man): HE (a&d), cleaved caspase-3 (b&e) and CD68 (c&f). In CN, fragmented apoptotic bodies, clustered under the chained migrated nuclei, are immunoreactive for cleaved caspase-3. In contrast, in the surgically removed normal colorectal mucosa, cleaved caspase-3-immunoreactive rounded nuclei are extruded onto the apical surface. CD68-positive macrophages located in the upper part of the lamina propria mucosae actively phagocytize apoptotic debris in the plump and amoeboid cytoplasm, while macrophages in the normal mucosa are round in shape and smaller in size.

#### 4. Ultrastructural features of CN

Buffered formalin-fixed biopsy tissue of CN (n=2) was dug out of paraffin blocks for the ultrastructural analysis [7]. After deparaffinization, tissue blocks were prepared with the conventional sequence. Ultrathin sections were stained with uranyl acetate and lead citrate. It is of note that ultrastructural morphology was satisfactory even after paraffin embedding. As illustrated in Figure 5, apoptotic nuclear debris was localized within the cytoplasm beneath the migrated nuclei of the surface-lining columnar cells. Abnormality of cytoskeletal filaments was scarcely recognized in the epithelial cytoplasm. Macrophages located in the uppermost part of the lamina propria phagocytized electron-dense globular apoptotic materials. IELs with scattered dense bodies were scattered among the columnar cells.



**Figure 5:** Ultrastructural features of CN with IBD-like symptoms (54-year-old man). (a) Low-powered views of transmission electron microscopy for CN. (b–d) High-powered views of transmission electron microscopy for CN. (a) The nuclei of the surface-lining columnar cells are migrated in chain to the middle part of the cells. In the uppermost part of the lamina propria mucosae, macrophages phagocytizing electron-dense apoptotic bodies (arrowheads) are clustered, and lymphocytes are also moderately infiltrated (bar=10  $\mu$ m). (b) An IEL is indicated by an arrowhead, and apoptotic debris (arrow) is seen among the epithelial cells. No aggregation or reorganization of cytoskeletal filaments is observed in the infranuclear cytoplasm (bar=5  $\mu$ m). (c) A close view of a macrophage observed in the uppermost



part of the lamina propria, actively phagocytizing electron-dense apoptotic nuclear debris (bar=5 µm). (d) The dense bodies are dispersed in the cytoplasm of the IEL among the surface-lining columnar cells (bar=2 µm).

## **5. Discussion**

Recent epidemiological studies has suggested that MC is more common than initially expected: one study identified MC in 10% of colorectal biopsy specimens from patients with non-bloody diarrhea, and in >20% of such patients are older than 70 years [3,4]. Reportedly, MC occurs more frequently in women than in men, and usually affects patients in the sixth and seventh decades [3,4]. In our analysis, however, all the cases of CC were men. In 15% of MC cases, a range of mild endoscopic abnormality was reported, such as patchy reddening/inflammation, pancolitis and featureless mucosa [5]. The etiology of MC remains unclarified, but oral medication has been suggested as a potential trigger: The drugs include lansoprazole, simvastatin, flutamide, ranitidine, carbanazepin, vinburninem paroxetine, sertraline, penicillin V, pivmecillinam, orlistate, tardyferon and Cyclo 3 Fort [8]. Autoimmune disorders may be associated with MC, in a range of 20-60% for both CC and LC [8].

We proposed CN as the third-type MC, microscopically characterized by chained migration of the nucleus in the colorectal surface-lining epithelium, in association with accelerated apoptosis of debris type beneath the nuclei [6]. Patients with CN were categorized into two clinical/endoscopic subtypes. One group manifested MC-like symptoms (watery non-bloody diarrhea) with minimal endoscopic abnormality. Another group is featured by IBD-like symptoms (occult/gross hematochezia, abdominal pain and/or mucin secretion) with patchy reddening of the colorectal mucosa with or without erosions/aphthae.

When compared with conventional MC (CC and LC), CN showed a male predominance, with a wide age range from 17 to 88 years (median: 63 years). Some patients received medication of PPIs as a potential trigger, as is so in conventional MC. In three cases of CN with MC-like features, diarrhea improved after discontinuation of PPIs. In four patients (one in MC-like type, and three in IBD-like type), CN occurred during or several months after chemotherapy against hematopoietic malignancies or colonic adenocarcinoma. Regarding autoimmune disorders, two patients of CN were clinically suspected of Behçet disease without intestinal ulceration, and a total of two patients suffered from IgA nephropathy or Sjögren syndrome. CN-type microscopic features were also seen in a remission state of ulcerative colitis. The candidate etiologic factors for provoking CN should thus include the medication of PPIs or anti-cancer drugs, the association of immune-related disorders and the altered cell turnover secondary to the treatment of IBD.

The pathogenesis of CN may be linked to the abnormality of epithelial apoptosis of debris type in the colorectal mucosa [6,7]. In the intestinal mucosa, the microbiotas may induce apoptosis of epithelial cells in balance with

adaptive immune homeostasis. Nakahashi-Oda et al. [8] recently documented that apoptotic epithelial cells negatively regulate the gut commensal bacteria-stimulated proliferation of regulatory T-cells, playing a central role in the maintenance of gut tissue homeostasis. Supposedly, apoptotic epithelial cells suppress the proliferation of regulatory T-cells for regulating mucosal homeostasis. Commensal enteric bacteria play an important role in autoimmune disorders [10]. In fact, dysbiosis of the gut normal flora has been observed in patients with autoimmune disorders, such as systemic lupus erythematosus [11]. In contrast, some microbiotas, especially *Clostridium IV* or XIV, may induce regulatory T-cells and inhibit the development of IBD [11]. The chained nuclear migration to the middle part of the surface-lining epithelial cells was quite unique and pathognomonic of CN, and easily recognizable in hematoxylin and eosin-stained biopsy specimens. Such microscopic appearance of CN was barely observed in CC and the control normal colorectal mucosa. The mechanisms of the nuclear migration remain uncertain. It is possible that abnormality of cytoskeletal proteins may impair localization of the nuclei that are normally anchored to the basal part of the columnar cells. However, negative electron microscopic findings were observed, as described above [7] (Figure 5). Under a low-powered magnification, pathologists may confuse CN with CC, because the eosinophilic cytoplasmic component beneath the migrated nuclei of the surface epithelial cells superficially looks like the collagen bands in CC. Therefore, in order to avoid inappropriate diagnosis, close-up microscopy under high magnification is inevitable.

The mode of apoptosis in normal gut mucosa has been analyzed microscopically. Sträter et al. [12] described two different patterns of enterocytic apoptosis: (a) apoptotic bodies (nuclear debris) were engulfed by adjacent epithelial cells, and (b) the apoptotic cells with only delicate morphological changes were extruded into the gut lumen. The engulfment pattern was observed predominantly in the crypt. The extrusion pattern was restricted to the luminal mucosal surface. Iwanaga et al. [13] showed unique features of apoptosis at the tip of small intestinal villi of the guinea pig. The apical cytoplasmic plates without containing nuclei were pinched off into the lumen, and the nuclei were engulfed by macrophages located in the upper part of the lamina propria mucosae. The fate of apoptotic nuclei of enterocytes may thus be dependent on the site in the mucosa and animal species. In the present study, the protrusion pattern of apoptosis was observed on the normal colorectal mucosa, just in accordance with the report by Sträter et al. [12]. CD68-positive macrophages actively phagocytizing apoptotic nuclear debris were more easily seen in CN than in the normal colorectal mucosa [6]. Abnormality of enterocytic apoptosis has so far been linked to a variety of intestinal disorders [14,15]. The chained nuclear migration in the surface-lining columnar cells, in association with fragmented nuclear debris beneath the nuclei, may represent an accelerated and altered apoptosis (switching from the extrusion type to debris type) in the diseased colorectal mucosa of CN [6].

IELs of CD8-positive cytotoxic T-cell type are known to promote epithelial apoptosis by secretion of cytotoxic molecules such as granzymes and perforin: Granzyme B accelerates apoptosis through both caspase-dependent and -independent pathways [16]. IELs also contribute to cryptal apoptosis and ulceration in active IBD [17]. In celiac

disease, increase of IELs expressing Fas ligand and perforin stirs up mucosal damage and epithelial apoptosis [18,19]. In acute human immunodeficiency virus infection, both apoptotic epithelial cells and IELs are increased [20]. In our analysis, cleaved caspase-3-immunoreactive fragmented apoptotic bodies were more frequently observed in IBD-like CN than in MC-like CN, but CB8-positive IELs and CD68-positive phagocytizing lamina propria macrophages were not significantly altered [6]. It is supposed that the accelerated apoptosis of debris type may be related to clinical association of occult/gross haematochezia. A recent study [21] showed in the UC rat model that microRNA-21-5p inhibited interleukin-6 receptor/signal transducer and activator of transcription signal-mediated pathway to decrease the level of inflammatory cytokines and apoptosis. Therefore, the authors implicated microRNA-21-5p as a candidate of therapeutic molecular target of UC in the human. The microRNA-21-5p-targeted therapy may be hopefully applicable to IBD-like CN.

The clinical and epidemiological features of MC have been established during the past 30 years [2,3]. We proposed a new variant of MC, "CN". It is of note that more than half (60%) of colorectal biopsy specimens sampled from patients with UC in a remission state showed microscopic appearance resembling CN [6]. The chained nuclear migration to the middle part of the surface-lining columnar cells may be correlated with the abnormality in apoptotic processes of the colorectal surface epithelial cells. Further clinicopathological studies are needed to evaluate whether CN can really be regarded as a distinct disease entity in the category of MC, and to settle unanswered questions in this perplexing condition, including the cause, pathophysiology, and optimal treatment.

### **Contributors**

MT and YT contributed equally to the present analysis. YT proposed a basic idea of CN. Both authors have participated sufficiently in the work to take public responsibility for appropriate portions of the content, cooperatively designed the study, and analyzed both light and electron microscopic features.

### **Funding**

The authors received no financial support for the research, authorship, and/or publication of the present article.

### **Competing Interests**

None declared.

### **Ethics Approval**

All the procedures were in accordance with the ethical standards of the responsible institutional committee on human experimentation and with the Helsinki Declaration of 1964 and later versions. The study was approved in December

2019 by the Ethics Committee for Clinical Research of Shimada Municipal Hospital, Shimada, Shizuoka, Japan (approval number R01-10).

### **Provenance and Peer Review**

Not commissioned; externally peer reviewed.

### **Patient Consent for Publication**

Written informed consent was obtained from the respective patients after selection of cases, including negative control cases.

### **Acknowledgments**

Tomohiko Hanaoka, M.D., Shinya Watanabe, M.D., Masahiro Matsushita, M.D., and Tadahiro Isono, M.D. at the Departments of Gastroenterology (TH, SW, MM) and Surgery (TI), Shimada Municipal Hospital, Shimada, Shizuoka, contributed to providing valuable clinical information. We cordially thank Naoki Ooishi, M.T., Kuniaki Muramatsu, M.T., and Kazuhiro Nakajima, M.T., Department of Diagnostic Pathology, Shimada Municipal Hospital, Shimada, Shizuoka, Japan. Kohei Watanabe, M.T., Special Reference Laboratories, Hamura, Tokyo, Japan, are also acknowledged for his support in the electron microscopic analysis.

### **References**

1. Choi EK, Appelman HD. Chronic colitis in biopsy samples. *Surg Pathol Clin* 10 (2017): 841-861.
2. Tysk C, Bohr J, Nyholm N, et al. Diagnosis and management of microscopic colitis. *World J Gastroenterol* 14(2008): 7280-7288.
3. Park T, Cave D, Marshall C. Microscopic colitis: a review of etiology, treatment and refractory disease. *World J Gastroenterol* 21 (2015): 8804-8810.
4. Baert F, Wouters K, D'Haens G, et al. Lymphocytic colitis: a distinct clinical entity? A clinicopathological confrontation of lymphocytic and collagenous colitis. *Gut* 45(1999): 375-381.
5. Williams JJ, Beck PL, Andrews CN, et al. Microscopic colitis: a common cause of diarrhoea in older adults. *Age Ageing* 39 (2010): 162-168.
6. Tachibana M, Hanaoka T, Watanabe S, et al. Colitis nucleomigrans: the third type of microscopic colitis (part 1). *Pathol Int* 70 (2020): 752-760.
7. Tachibana M, Tsutsumi Y. Colitis nucleomigrans: the third type of microscopic colitis (part 2). An ultrastructural study. *Pathol Int* 70 (2020): 761-766.
8. Olesen M, Eriksson S, Bohr J, et al. Microscopic colitis: a common diarrhoeal disease. An epidemiological study in Orebro, Sweden, 1993-1998. *Gut* 53 (2004): 346-350.

9. Nakahashi-Oda C, Udayanga KGS, Nakamura Y, et al. Apoptotic epithelial cells control the abundance of Treg cells at barrier surfaces. *Nat Immunol* 17 (2016): 441-450.
10. Bach JF. The hygiene hypothesis in autoimmunity: the role of pathogens and commensals. *Nat Rev Immunol* 18 (2018): 105-120.
11. Honda K, Littman DR. The microbiota in adaptive immune homeostasis and disease. *Nature* 535 (2016): 75-84.
12. Sträter J, Koretz K, Günthert AR, et al. In situ detection of enterocytic apoptosis in normal colonic mucosa and in familial adenomatous polyposis. *Gut* 37 (1995): 819-825.
13. Iwanaga T, Han H, Adachi K, et al. A novel mechanism for disposing of effete epithelial cells in the small intestine of guinea pigs. *Gastroenterology* 105 (1993): 1089-1097.
14. Ramachandran A, Madesh M, Balasubramanian KA. Apoptosis in the intestinal epithelium: its relevance in normal and pathophysiological conditions. *J Gastroenterol Hepatol* 15 (2000): 109-120.
15. Negroni A, Cucchiara S, Stronati L. Apoptosis, necrosis, and necroptosis in the gut and intestinal homeostasis. *Mediators Inflamm* 2015 (2015): 250762.
16. Henkart PA. Lymphocyte-mediated cytotoxicity: two pathways and multiple effector molecules. *Immunity* 1 (1994): 343-346.
17. Mitomi H, Ohkura Y, Yokoyama K, et al. Contribution of TIA-1+ and granzyme B+ cytotoxic T lymphocytes to cryptal apoptosis and ulceration in active inflammatory bowel disease. *Pathol Res Pract* 203 (2007): 717-723.
18. Ciccocioppo R, Di Sabatino A, Parroni R, et al. Cytolytic mechanisms of intraepithelial lymphocytes in coeliac disease (CoD). *Clin Exp Immunol* 120 (2000): 235-240.
19. Ciccocioppo R, Di Sabatino A, Parroni R, et al. Increased enterocyte apoptosis and Fas-Fas ligand system in celiac disease. *Am J Clin Pathol* 115 (2001): 494-503.
20. Epple HJ, Allers K, Tröger H, et al. Acute HIV infection induces mucosal infiltration with CD4+ and CD8+ T cells, epithelial apoptosis, and a mucosal barrier defect. *Gastroenterology* 139 (2010): 1289-1300.
21. Lu X, Yu Y, Tan S. The role of the miR-21-5p-mediated inflammatory pathway in ulcerative colitis. *Exp Ther Med* 19 (2020): 981-989.

**Citation:** Mitsuhiro Tachibana, Yutaka Tsutsumi. Colitis Nucleomigrans: A Proposal for The Third Type of Microscopic Colitis. *Archives of Clinical and Medical Case Reports* 5 (2021): 342-354.



This article is an open access article distributed under the terms and conditions of the [Creative Commons Attribution \(CC-BY\) license 4.0](https://creativecommons.org/licenses/by/4.0/)